

Surface Characterization of *Escherichia Coli*-Imprinted Polymers Using Confocal Raman Microscopy

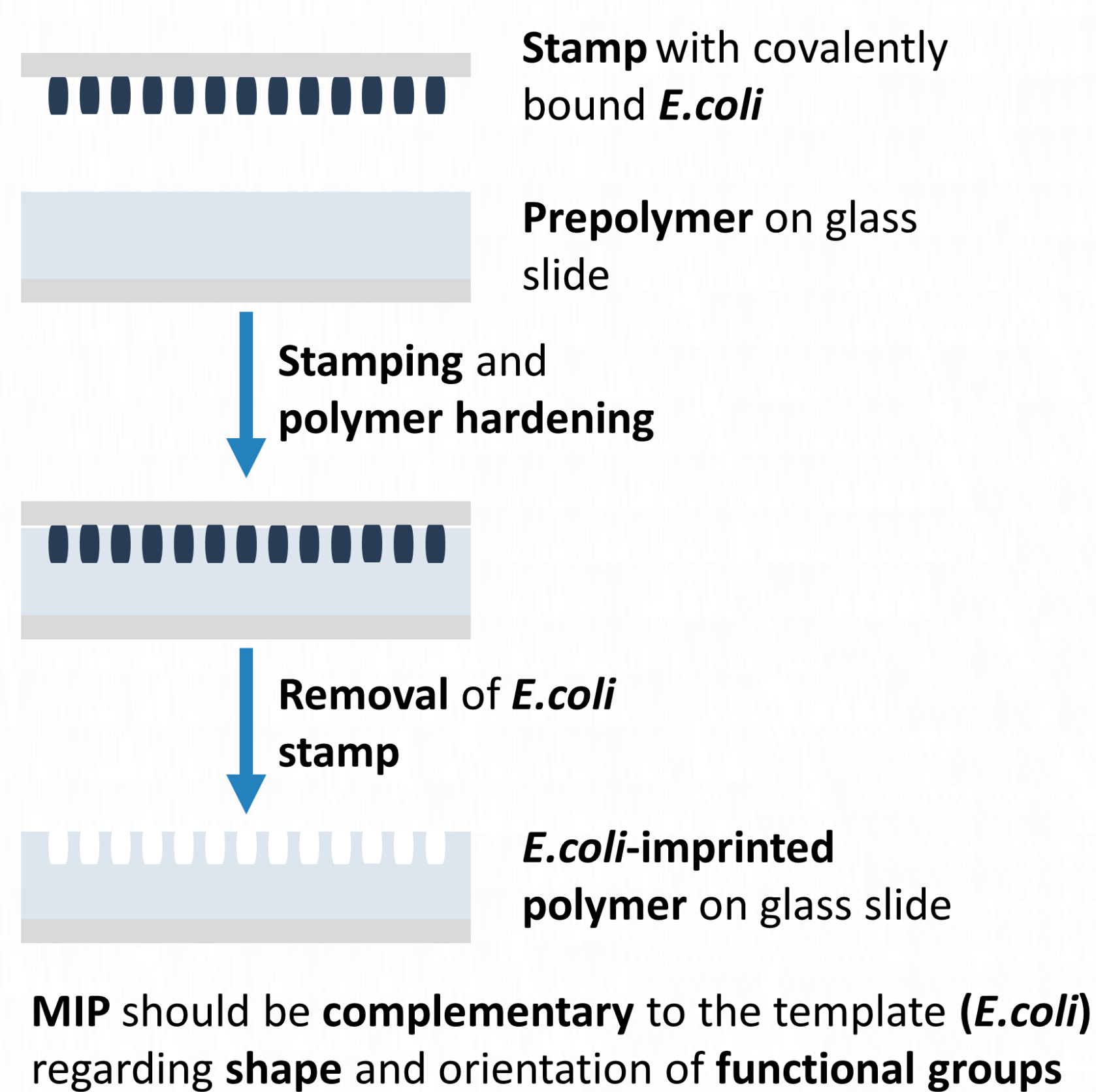
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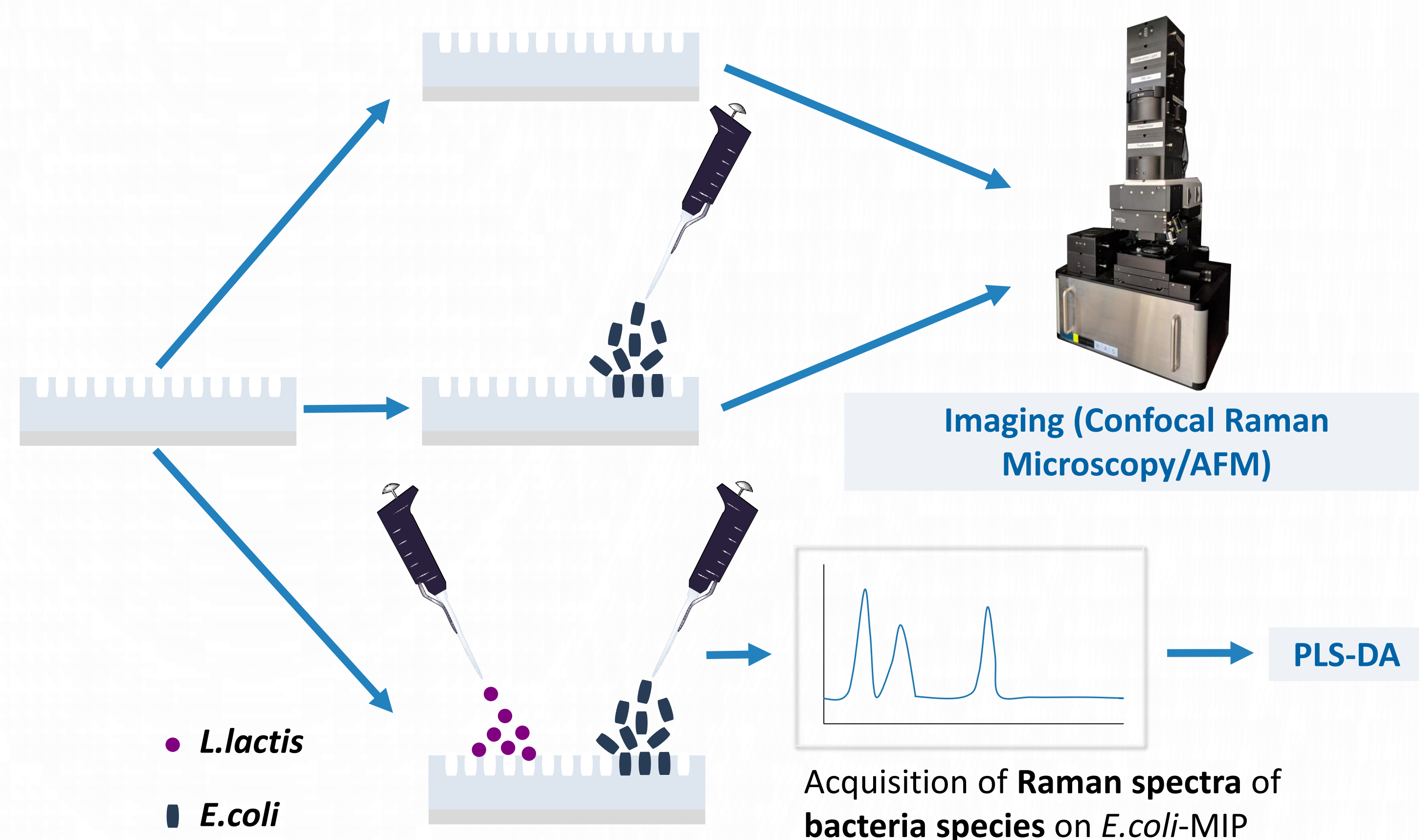
Introduction

The Gram-negative bacterium *Escherichia coli* (*E. coli*) is considered an **indicator of hygiene** in food and water. Therefore, a variety of methods has been established for the **detection, identification and quantification** of this microorganism, most of which require growing the bacteria. In addition, available techniques such as flow cytometry depend on trained staff with substantial **expertise** (1). However, it is possible to generate *E. coli*-sensitive sensors based on ***E. coli*-selective molecularly imprinted polymers (MIPs)** as receptors on Quartz Crystal Microbalances (QCMs) for **direct detection** of the microorganism in water (2). In order to render the synthesis of the MIPs reproducible and to assess success of the imprinting, different techniques have been established to **characterize MIPs**, including Atomic Force and Optical Microscopy. Both provide topological information but **no information** about **chemical composition** of the surface. We employed Confocal Raman Microscopy for **optical and chemical** surface characterization of *E. coli*-MIPs (poly(styrene-co-divinylbenzene)) and assessment of success of the **imprinting procedure**. Furthermore, we combined Confocal Raman Microscopy with **Partial Least Squares Discriminant Analysis (PLS-DA)** to distinguish different bacteria species on the *E. coli*-MIP for further **selectivity studies**.

E. coli-Imprinted Polymers



Confocal Raman Microscopy and PLS-DA



Results

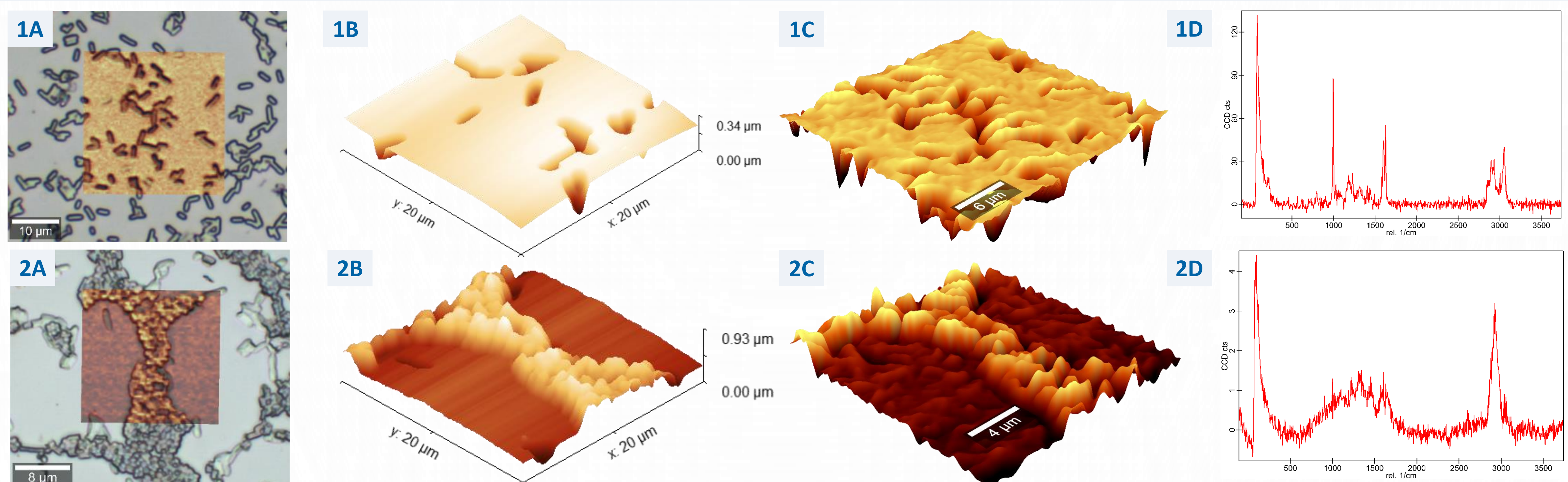


Fig. 1-2: AFM and Raman images of untreated (1B, 1C) and *E. coli*-treated MIPs (2B, 2C). AFM measurements **confirmed the imprinting of *E. coli*** on poly(styrene-co-divinylbenzene) (1B) and the presence of *E. coli* on the *E. coli*-treated MIP (2B). **Overlaying of images of the Raman intensity at 2908 cm⁻¹** (using WiTec Raman TV) obtained from the **untreated (1C) and *E. coli*-treated MIP (2C)** with the corresponding **white light images (1A and 2A)** showed that **Confocal Raman Microscopy** allowed for straightforward **differentiation between imprints, polymer and *E. coli***. The accuracy of this differentiation was also confirmed by AFM scans on the exact same sample area in the case of *E. coli*-treated MIP (2B). **WiTec True Component Analysis** of Raman image scans of *E. coli*-treated MIP yielded **2 components: *E. coli* attached to poly(styrene-co-divinylbenzene) (1D) and plain poly(styrene-co-divinylbenzene)**. Subtraction of the plain polymer component from the other one yielded a spectrum similar to a clean *E. coli* spectrum (2D). This confirmed the presence and location of *E. coli* on the MIP suggested by the AFM and simple Raman intensity images at 2908 cm⁻¹.

Raman imaging parameters: 532 nm laser wavelength, 8 mW laser power, 0.1 s integration time per spectrum, all images and spectra acquired on a WiTec alpha 300 system

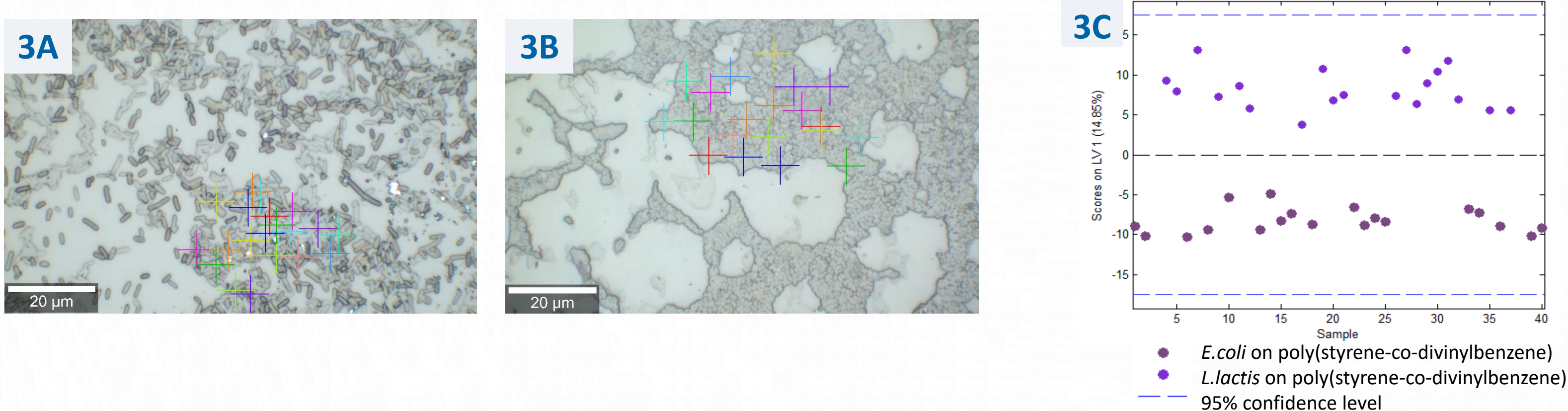


Fig. 3: Single spectra of *E. coli* and *L. lactis* were acquired on *E. coli*-imprinted poly(styrene-co-divinylbenzene) at the locations indicated in 3A (*E. coli*) and 3B (*L. lactis*). The spectra were preprocessed (baseline correction, normalization and autoscaling) and a **PLS-DA model** with one latent variable was established to successfully **differentiate** between the two **bacteria species** on the polymer surface (3C).

Raman parameters for single spectra: 532 nm laser wavelength, 8mW laser power, 3x20s integration time

Conclusion

Confocal Raman Microscopy was successfully used for the assessment of imprinting success in *E. coli*-imprinted poly(styrene-co-divinylbenzene). Furthermore, different bacteria species, *E. coli* and *L. lactis*, could be differentiated on the surface of the MIP using a combination of Confocal Raman Microscopy and Partial Least Squares Discriminant Analysis, which provides a basis for further selectivity studies of the polymer.

References

- Davey, Hazel M. and Kell, Douglas B. Flow Cytometry and Cell Sorting of Heterogeneous Microbial Populations: The Importance of Single-Cell Analyses. Microbiological Reviews. 1996, Vol. 60, pp. 641-696.
- Poller, Anna-Maria, et al. Surface Imprints: Advantageous Application of Ready2use Materials. ACS Applied Materials and Interfaces. 2017, Vol. 9, pp. 1129-1135.